

International Journal of Advances in Engineering and Management (IJAEM) Volume 3, Issue 9 Sep 2021, pp: 624-636 www.ijaem.net ISSN: 2395-5252

# **Utilization of Solid Waste for Treating** Leachate

### Karthik Boppana

Submitted: 01-09-2021

Revised: 09-09-2021

Accepted: 12-09-2021

### ABSTRACT

The adsorption process is widely used for the removal of heavy metals from leachate because of its low cost, availability and eco-friendly nature. Both commercial adsorbents and bio-adsorbents are used for the removal of heavy metals from leachate, with high removal capacity. Hence in many of the landfills, adsorption is one of the very commonly used method. It is therefore very important that the adsorbents used are economical; considering the fact that these the leachate has no other use attached to it. Furthermore if the adsorbents are solid wastes or from such sources, it would be more effective and efficient considering the whole scenario.

Key words: Leachate, heavy metal, adsorption, removal rate

### I. INTRODUCTION

Leachate is the liquid formed when waste breaks down in the landfill and water filters through that waste. This liquid is highly toxic and can pollute the land, groundwater and waterways. This problem could be worsened in the case of a landfill leachate, which is the liquid that exists as part of rainwater entering the landfill but is also due to the natural decomposition of organic material along with other liquids and chemicals that have been discarded. Rainwater passes through the waste in the landfill and if the landfill is not properly lined or the leachate is not properly managed, it is at great risk of mixing with the groundwater near the site or any water body, in the vicinity, especially in the lower gradient.

The pollutant composition of the leachate is important so that appropriate treatment systems could be installed for reducing or eliminating them. Leachates are composed of organic and inorganic substances. Organic substances consist of microorganisms, their metabolic products and materials from living organisms which are undergoing decay. Inorganic pollutants in the leachate consist of ammonium, phosphorous,

\_\_\_\_\_ sulphate and metals. Along with the substances mentioned above leachate contains many others that are undesirable because of their negative effect on the environment and human life.

-----

Inorganic substances can have an impact on turbidity and deposits on pipes (Iron), increase the hardness of water (Calcium & Magnesium). Organic substances have an impact on colour, odour and taste of water. Nutrients such as ammonium and phosphorus contribute to the eutrophication of receiving waters which can lead to algae blooms. An important part of maintaining a landfill is the managing of the leachate to prevent pollution into surrounding ground and surface waters.

Since the discharge of the leachate into the ground and thus would find the way into ground water and nearby watersources such as lakes or rivers, if any change in altitude. The leachate would consist of wastes from industry and other sources containing organic and inorganic pollutants. Manyatimes the leachate consists of high levels of heavy metals which should be removed it so that it wouldn't affect the water sources by contamination. Often these heavy metals are toxic/or carcinogenicwhich are harmful to both humans and other living species.

Heavy metals pollution has raised serious environmental concerns worldwide because bioaccumulation of these elements beyond the tolerance thresholds of living organisms pose long term risk to the earth's ecosystem (Voegelinet al., 2003; Sparks, 2005). The main flows of heavy metals to the environment are from industrial and municipal wastes, both of which contained a variety of toxic heavy metals.

Heavy metals in leachate from landfills have been extensively studied and monitored (Yong, 2001; Selim and Sparks, 2001). The major part of the metals is retained in the landfill. As a consequence, it must be expected that leaching of heavy metals from the landfills will continue for a long time (Freeze and Cherry, 1979; Fetter, 2001;



Selim and Sparks, 2001; Yong, 2001). It can take years before groundwater pollution reveals itself and chemicals in the leachates often react synergistically and often in unanticipated ways to affect the ecosystem (Lee and Sheehan, 1996).

The heavy of most concern from industries include Lead(Pb), Zinc(Zn), Copper(Cu), Arsenic(As), Cadmium(Cd), Chromium(Cr), Nickel(Ni). They originate from sources such as metal complex dyes, pesticides, fertilizers, fixing agents etc.

Though there are many treatment technologies available for aiding in the heavy metal removal one very effect method most commonly used is adsorption.

### **II. OBJECTIVE**

In this paper proper understanding of different solid wastes which could act as an potential adsorbent has been made. Out of the total potentialadsorbents based on the source they have been classified namely as natural adsorbents, agriculturalwaste, industrial solid waste and plastic waste.

And for all the above-mentionedpotential adsorbents, the assessment of heavy metal adsorption has been made referring to different factors which play a important role in the rate of adsorption and as well the quantity of adsorption. These factors include the influence of adsorbent dosage, effect of ph, effect of contact time etc.

And based on these factors the most suitable adsorbent and their optimum conditions have been identified.

### III. MATERIALS AND METHODS ADSORPTION

Adsorption relies on the physical and chemical interactions between heavy metal ions and adsorbents.

#### Usage of Solid Waste as Adsorbents

For the treatment of leachate adsorbents of high cost might not be suitable as because the main purpose is only stopthe leachate fromcontaminating the groundwater and nearby water sources if any. So, the usage of the Solid waste would be very useful and efficient solution.

Some of the solid waste for adsorption could be mainly split into different types based on their source namely natural adsorbents, industrial solid waste, inorganic solid wate, municipal waste, plastic waste.

The natural adsorbents include vegetable fibres, rice husk, straw, sawdust, peat, hay, kapok, certain kinds of wood etc.

The industrial solid waste include NHISW, red mud, activated carbon, alkali modified fly ash, food industry waste, granite and marble industry waste, class 4 bricks and brick powder, raw clay and broken clay-brick waste.

Inorganicsolid waste includes air-cooled blast furnace slag, water quenched blast furnace slag, steel furnace slag, coal fly ash, coal bottom ash. water treatment sludge, red mud.

The plastic wastes which could be used for adsorption include polypropylene waste, low density polyethylene (LDPE) waste, modified PET fiber, polyacrylic acid-polyvinylidene fluoride (PVDF) blende polymer adsorbent etc

### **IV. THE RESULTS**

### A. Soil Type

The adsorption results of the soils of different types and their ability to adsorb heavy metals

1) **For Copper** – The maximum adsorption occurs by using clay soil as an absorbent. Theminimum adsorption occurs by using silty soil as an absorbent.

2)For Zinc- The maximum adsorption occurs by using claysoil as an absorbent The minimum adsorption occurs by using silty soil as an absorbent.

3) For Manganese – The maximum adsorption occurs by using claysoil as an absorbent The maximum adsorption occurs by using clay soilas adsorbent.

4)**For Chromium** – The maximum adsorption occurs by using clayey soil as an absorbent The minimum adsorption occurs by using siltysoil as an absorbent.

Among all the soils, the maximum adsorption occurs in the clayey soil with zinc. The minimum adsorption occurs in the silty soil with copper.



### **B.** Natural organicadsorbents

The below table shows a list of different types of natural adsorbents which could be use and their removal rate

Adsorbent	Heavy metals	Remarks	Source
Palm Kernel Shell	Chromium, Lead, Zinc	Highest contact	Baby and Hussein, 2020
	and Cadmium	time 120 min	
Palm Kernel Shell	Cadmium	Highest contact	Faisal et al.,2019
		time 150 min,	
		decreases at	
		180min	
Pomegranate Peel	Nickle	Sharply	Elsayed et al.,2020
		increased	
		during the first	
		30min;	
		gradually	
		achieved the	
		equilibrium in	
		150 min	
Kenaf Fibre	Iron, Manganese, Zinc,	The contact	Saeed et al.,2020
	Arsenic, Copper, Nickle	time will	
		eventually	
		reach a	
		maximum	
		value at a	
		certain point	
		and remain	
Maria		constant	D
Mango Leaf	Chromium and Iron	Adsorption	Duraisamy et al.,2020
		takes place at 120min of	
		interaction time	
Coffee Shell	Lead	In the first	Junair et al.,2019
Conce Shell	Lead	30min until	Julian et al.,2019
		min. 90 the	
		adsorption rate	
		is slow;	
		however from	
		minute 90 until	
		min. 150	
		ultimately the	
		adsorption	
		equilibrium	
		occurs	
Chestnut Shell	Chromium	An increase in	Singh et al.,2020
		adsorption was	
		seen at initial	
		60-300 min	
		thereafter	
		remained	
		constant	
Jackfruit Peel	Lead and Chromium	Range of	Ibrahim et al.,2020
		between	
		15minutes to	
		24hours.	
		Adsorption was	
		rapid during the	



		first 1 hour of contact but gradually decreases up to the point where equilibrium is achieved.
Oil Palm Ash	Manganese	Within 80min Chowdhury et al.,2011 the system reached equilibrium

Table 1 – Credits Journal of Ecological Engineering 2021, 22(3), 249-265

The rate of adsorption depends on a number of factors namely the effect of ph, adsorbent dosage and contact time

### Effect of Ph

For the set of natural adsorbents, the effect of pH on Heavy metal adsorption has also been specified

Adsorbent	Heavy metal	pH range	Optimum	Adsorption	Source
			pH value	capacity	
Pomegranate	Nickle	4 to 9	9	98%	Elsayed et al.,
peel					2020
Kenaf Fibre	Iron,	3 to 11	7	Between 5% to	Saeed et al .,
	Manganese,			30%	2020
	Zinc, Arsenic,				
	Copper, Nickle				
Mango Leaf	Chromium and	2 to 10	8	99% to 99.5%	Duraisamy et
	Iron				al., 2020
Palm Kernel	Chromium,	2 to 6	6	60% to 80%	Baby and
Shell	Lead, Zinc and				Hussein, 2020
	Cadmium				
Chestnut Shell	Chromium	2 to 12	7	78%	Singh et
					al.,2020
Banana Peel	Copper, Nickle	0.6 to 7.4	5.7 to 7.4	40%, 51% and	Thuan et al.,
	and Lead			54%	2017
Jackfruit Peel	Lead and	4 to 9	7	50% to 90%	Ibrahim et al.,
	Cadmium				2020
Pistachio Hull	Nickle	2 to 10	6	60% to 90%	Beidokhti et al.,
					2019
Oil Palm Ash	Copper	2 to 8	8	50% to 94%	Chowdary et al.,
					2011
Oil Palm Shells	Nickle, Lead	3 to 10	8	Up to 70%	Rahman et al.,
	and Chromium				2014

Table 2 – Credits Journal of Ecological Engineering 2021, 22(3), 249-265

### Influence of Adsorbent Dosage

The influence of adsorbent dosage on the removal of heavy metal has been specified

Adsorbent	Adsorbent dose	Heavy metals	Removal capacity	Source	
Mango Leaf	20 to 100mg/l	Chromium and Iron	Chromium: from 93.4% to 99.6% Iron: from 89.4% to 99.4%	Duraisamy al.,2020	et



**International Journal of Advances in Engineering and Management (IJAEM)** Volume 3, Issue 9 Sep 2021, pp: 624-636 www.ijaem.net ISSN: 2395-5252

Chestnut Peels	0.2 to 1.0 g	Chromium	60% to 79%	Singh et al., 2020
Banana Peels	0.9 to 2.4 g/l	Copper, Nickle and Lead	Copper: 40%, Nickle: 51% and Lead: 54%	Thuan et al.,2017
Pistachio Hull	5 to 30 g/l	Nickle	66% to 76%	Beidokhti et al.,2019

Table 3 – Credits Journal of Ecological Engineering 2021, 22(3), 249-265

### Effect of contact time

Variation of contact time for diverse adsorption systems

Adsorbent	Metal ions	Conditions	Maximum adsorption
			capacity (mg/g)
Sweet Potato Peels	Pb(II),	Contact time:0-60 min	200.91 mg/g for Pb(II)
	Cd(II)	pH: 2-12	125 mg/g for Cd(II)
		Adsorption dosage: 0.5g	
		Temperature: 30-80 C	
		Initial concentration: 10-	
		80 mg/L	
Musa paradisiaca peels	Pb(II),	Contact time:15-90 min	10 mg/g for both metal
	Cd(II)	pH: 5-8	ions
		Adsorption dosage: 0.1-	
		0.7 g	
		Temperature: 30-60 C	
		Initial concentration: 50-	
		200 mg/L	
Activated carbon from	Cr(VI),	Contact time:0-300 min	144.93 mg/g for Cr(VI)
molasses	Pb(II),	pH: 2-11	303.03 mg/g for Pb(II)
	Cu(II)	Initial concentration: 25	526.32 mg/g for Cu(II)
		mg/L of Cu(II), 23.4	
		mg/L of Pb(II) and 24.3	
		mg/L of Cr(VI)	
Grounnut(Arachis	Pb(II),	Contact time:0-300 min	94.07 mg/g for Pb(II)
hypogaen) shell	Cd(II),	pH: 1-6	104.71 mg/g for Cd(II)
	Zn(II)	Adsorption dosage: 0.1-	86.13 mg/g for Zn(II)
		2g	
		Temperature: 20-45 C	
		Initial concentration: 10-	
		100 mg/L	

Table 4 – Credits J Mater Res Technol, 2020, 9(5), 10235-10253

The adsorption values in the case of oil palm constituents and derivates along with their influence with change in pH, contact time, dosage has been specified

### Optimum values of affecting factors for heavy metals adsorption using Oil Palm AC

Oil Pal	m AC	Heavy metals	Factor values)	affecting adsorp	Source	
			pH value	Contact time	AC dose	
Palm	Kernel	Lead	4	60 min	1.5 g	Baby and
Shell		Chromium	6	60 min	1.5 g	Hussein., 2019
		Cadmium	6	90 min	2.0 g	
		Zinc	6	120 min	2.0 g	
Oil	Palm	Manganese	7	60 min	N/A	Alothman et al.,



Leaves	Lead	6	60 min		2019
	Cobalt	7	60 min	-	
Oil Palm Ash	Manganese	7	80 min	N/A	Chowdhury et
					al., 2011
Palm Kernel	Chromium	6	120 min	0.25 g	Baby and
Shell	Lead				Hussien, 2020
	Zinc				
	Cadmium				
Palm Fruit	Lead	5	120 min	N/A	Oola and Ong,
Fibre					2019

Table 5 – Credits Journal of Ecological Engineering 2021, 22(3), 249-265

### **Adsorption of Copper**

Maximum capacities for Adsorption of copper by different adsorbents

-49.0AnaerobicallyDigested SludgeGould (1978)-1.40Kaolin ClayFarah et al. (1980)-2.54Illite Clay23.3Montmorillonite clay-4.44-Treated bagaseKumar and Dara (13.46-Treated bagaseKumar and Dara (13.69-Treated Acacia Bark-3.69-Treated Techtona Bark-1.53-Fly-ashPanday et al. (1985)3.58-Rice HullsSuemitsu et al. (1977.88-Dyestuff-Treated (Red) Rice Hulls-7.00-Dyestuff-Treated (Yellow) Rice Hulls-7.3-Tea LeavesTan and Abd. R (1988)-14.0Amorphous Iron HydroxideMustafa and Haq (1-9.22Activated CarbonFerro-Gracia et al.0.438-ChitinGonzalez-Davila Millero (1990)-6.89Aspergillus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzae14.018.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoglocaramigera-1.89-Oil-Palm Fibre1.98Natural Oil-Palm Fibre1.98Natural Oil-Palm Fibre1.98Natural Oil-Palm Fibre1.98Natural Oil-Palm Fibre1.98Natural Oil-Palm Fibre	
-         1.40         Kaolin Clay         Farrah et al. (1980)           -         2.54         Illite Clay         -           4.44         -         Treated bagasse         Kumar and Dara (1           3.46         -         Treated Acacia Bark         -           3.08         -         Treated Acacia Bark         -           3.69         -         Treated Techtona Bark         -           1.53         -         Fly-ash         Panday et al. (1985)           3.58         -         Rice Hulls         Suemitsu et al. (1987)           7.88         -         Dyestuff-Treated (Red)         Rice Hulls           7.00         -         Dyestuff-Treated(Yellow)         Rice Hulls           7.10         -         Tea Leaves         Tan and Abd. R           -         14.0         Amorphous Iron Hydroxide         Mustafa and Haq (1988)           -         14.0         Amorphous Iron Hydroxide         Mustafa and Haq (1990)           -         6.89         Aspergillus oryzae         Huang et al (1991)           -         6.06         Rhizopus oryzae         Huang et al (1991)           -         6.06         Rhizopus oryzae         Ituan et al. (1992)           -	enetelli
-         2.54         Illite Clay           -         23.3         Montmorillonite clay           4.44         -         Treated bagasse         Kumar and Dara (1           3.46         -         Treated Acacia Bark         3.08           3.08         -         Treated Laurel Bark         3.308           1.53         -         Fly-ash         Panday et al. (1985)           3.58         -         Rice Hulls         Suemitsu et al. (1987)           7.88         -         Dyestuff-Treated (Red) Rice Hulls         Suemitsu et al. (1987)           7.00         -         Dyestuff-Treated (Yellow) Rice Hulls         Tan and Abd. R (1988)           27.3         -         Tea Leaves         Tan and Abd. R (1988)           -         14.0         Amorphous Iron Hydroxide         Mustafa and Haq (1990)           -         9.22         Activated Carbon         Ferro-Gracia et al.           0.438         -         Chitin         Gonzalez-Davila           Millero (1990)         -         6.89         Aspergillus oryzae         Huang et al (1991)           31.8         -         Treated Aspergillus oryzae         14.092)         1990           -         Sludge Solid         Tien and Huang (19	
-       23.3       Montmorillonite clay         4.44       -       Treated bagasse       Kumar and Dara (1         3.46       -       Treated Acacia Bark	)
4.44       -       Treated bagasse       Kumar and Dara (1         3.46       -       Treated Acacia Bark	
3.46       -       Treated Acacia Bark         3.08       -       Treated Laurel Bark         3.69       -       Treated Techtona Bark         1.53       -       Fly-ash       Panday et al. (1985)         3.58       -       Rice Hulls       Suemitsu et al. (1985)         7.88       -       Dyestuff-Treated (Red) Rice Hulls       Suemitsu et al. (1985)         7.00       -       Dyestuff-Treated (Yellow) Rice Hulls       Tan and Abd. R (1988)         27.3       -       Tea Leaves       Tan and Abd. R (1988)         -       9.22       Activated Carbon       Ferro-Gracia et al.         0.438       -       Chitin       Gonzalez-Davila Millero (1990)         -       6.89       Aspergillus oryzae       Huang et al (1991)         -       6.06       Rhizopus oryzae       14.0         13.8       -       Treated Aspergillus oryzae       14.0         -       6.06       Rhizopus oryzae       14.0         -       5.9       Aspergillus oryzae       14.0         -       6.06       Rhizopus oryzae       10.1         -       13.8       -       Treated Aspergillus oryzae         -       Sludge Solid       Tien and Huang (19	
3.08       -       Treated Laurel Bark         3.69       -       Treated Techtona Bark         1.53       -       Fly-ash       Panday et al. (1985)         3.58       -       Rice Hulls       Suemitsu et al. (1987)         7.88       -       Dyestuff-Treated (Red) Rice Hulls       Suemitsu et al. (1987)         7.00       -       Dyestuff-Treated (Yellow) Rice Hulls       Tan and Abd. R (1988)         27.3       -       Tea Leaves       Tan and Abd. R (1988)         -       14.0       Amorphous Iron Hydroxide       Mustafa and Haq (1988)         -       9.22       Activated Carbon       Ferro-Gracia et al.         0.438       -       Chitin       Gonzalez-Davila Millero (1990)         -       6.89       Aspergillus oryzae       Huang et al (1991)         -       6.06       Rhizopus oryzae       11.0         13.8       -       Treated Aspergillus oryzae       11.091)         31.8       -       Sludge Solid       Tien and Huang (192)         29.0       -       Zoogloearamigera       1.0         1.89       -       Oil-Palm Fibres       Low et al. (192)         29.0       -       Zoogloearamigera       1.0         1.	1982)
3.69       -       Treated Techtona Bark         1.53       -       Fly-ash       Panday et al. (1985)         3.58       -       Rice Hulls       Suemitsu et al. (1987)         7.88       -       Dyestuff-Treated (Red) Rice Hulls       Suemitsu et al. (1987)         7.00       -       Dyestuff-Treated (Yellow) Rice Hulls       Tan and Abd. R (1988)         27.3       -       Tea Leaves       Tan and Abd. R (1988)         -       14.0       Amorphous Iron Hydroxide       Mustafa and Haq (1988)         -       9.22       Activated Carbon       Ferro-Gracia et al.         0.438       -       Chitin       Gonzalez-Davila Millero (1990)         -       6.89       Aspergillus oryzae       Huang et al (1991)         -       6.06       Rhizopus oryzae       14.0 (1991)         31.8       -       Treated Aspergillus oryzae       1991)         31.8       -       Sludge Solid       Tien and Huang (1992)         29.0       -       Zoogloearamigera       1.0 (1993	
1.53-Fly-ashPanday et al. (19853.58-Rice HullsSuemitsu et al. (1987)7.88-Dyestuff-Treated (Red) Rice HullsRice Hulls7.00-Dyestuff-Treated(Yellow) Rice Hulls27.3-Tea LeavesTan and Abd. R (1988)-14.0Amorphous Iron HydroxideMustafa and Haq (1988)-14.0Amorphous Iron HydroxideMustafa and Haq (1988)-0.438-ChitinGonzalez-Davila Millero (1990)-6.89Aspergillus oryzaeHuang et al (1991)-6.06Rhizopus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzaeHuang et al (1991)31.8-Sludge SolidTien and Huang (1992)29.0-ZoogloearamigeraI.89-15.9Dye-Treated Oil-Palm FibreLow et al. (1993)-15.9Dye-Treated G. lucidumNagendra et al. (19-64.5Treated G. lucidumNagendra et al. (19-10.1Treated A. nigerNagendra et al. (19	
3.58-Rice HullsSuemitsu et al. (1997.88-Dyestuff-Treated (Red) Rice Hulls(Red) Rice Hulls7.00-Dyestuff-Treated(Yellow) Rice Hulls27.3-Tea LeavesTan and Abd. R (1988)-14.0Amorphous Iron HydroxideMustafa and Haq (1 (1988)-9.22Activated CarbonFerro-Gracia et al.0.438-ChitinGonzalez-Davila Millero (1990)-6.89Aspergillus oryzaeHuang et al (1991)-6.06Rhizopus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzae14.035.7-Sludge SolidTien and Huang (19 42.929.0-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19-10.1Treated A. niger	
7.88-Dyestuff-Treated (Red) Rice Hulls7.00-Dyestuff-Treated(Yellow) Rice Hulls27.3-Tea Leaves27.3-Tea Leaves-14.0Amorphous Iron Hydroxide-9.22Activated Carbon-9.22Activated Carbon0.438-Chitin0.438-Chitin0.438-Chitin0.438-Chitin0.438-Chitin13.8-Aspergillus oryzae13.8-Aspergillus oryzae13.8-Sludge Solid35.7-Sludge Solid42.9-Chlorella vulgarise1.89-Oil-Palm Fibres-1.98Natural Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidum-10.1Treated A. niger	5)
Rice Hulls7.00-Dyestuff-Treated(Yellow) Rice Hulls27.3-Tea LeavesTan and Abd. R (1988)-14.0Amorphous Iron HydroxideMustafa and Haq (1-9.22Activated CarbonFerro-Gracia et al.0.438-ChitinGonzalez-Davila Millero (1990)-6.89Aspergillus oryzaeHuang et al (1991)-6.06Rhizopus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzaeHuang et al (1991)31.8-Sludge SolidTien and Huang (192)29.0-CoogloearamigeraLow et al. (1992)29.0-Oil-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19-10.1Treated A. nigerNagendra et al. (19	
Rice Hulls7.00-Dyestuff-Treated(Yellow) Rice Hulls27.3-Tea Leaves14.0Amorphous Iron HydroxideMustafa and Haq (1988)-14.0Amorphous Iron HydroxideMustafa and Haq (1988)-9.22Activated CarbonFerro-Gracia et al.0.438-ChitinGonzalez-Davila Millero (1990)-6.89Aspergillus oryzaeHuang et al (1991)-6.06Rhizopus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzaeHuang et al (1991)31.8-Sludge SolidTien and Huang (192)29.0-CoogloearamigeraAksu et al. (1992)29.0-ZoogloearamigeraLow et al. (1993)-15.9Dye-Treated Oil-Palm FibreLow et al. (1993)-1.98Natural Oil-Palm Fibre28.5G. lucidumNagendra et al. (19-10.1Treated A. niger-	
Rice Hulls27.3-Tea LeavesTan and Abd. R (1988)-14.0Amorphous Iron HydroxideMustafa and Haq (1-9.22Activated CarbonFerro-Gracia et al.0.438-ChitinGonzalez-Davila Millero (1990)-6.89Aspergillus oryzaeHuang et al (1991)-6.06Rhizopus oryzaeHuang et al (1991)31.8-Aspergillus oryzaeHuang et al (1991)31.8-Sludge SolidTien and Huang (1942.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-15.9Dye-Treated Oil-Palm Fibre1.98Natural Oil-Palm Fibre28.5G. lucidumNagendra et al. (19-64.5Treated A. niger.	
Rice Hulls27.3-Tea LeavesTan and Abd. R (1988)-14.0Amorphous Iron HydroxideMustafa and Haq (1988)-9.22Activated CarbonFerro-Gracia et al.0.438-ChitinGonzalez-Davila Millero (1990)-6.89Aspergillus oryzaeHuang et al (1991)-6.06Rhizopus oryzaeHuang et al (1991)31.8-Aspergillus oryzaeHuang et al (1991)31.8-Sludge SolidTien and Huang (1942.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-0il-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19-10.1Treated A. nigerNagendra et al. (19	
Image: constraint of the systemImage: constraint of the systemImage: constraint of the system-14.0Amorphous Iron HydroxideMustafa and Haq (1)-9.22Activated CarbonFerro-Gracia et al. (1)0.438-ChitinGonzalez-Davila Millero (1990)-6.89Aspergillus oryzaeHuang et al (1991)-6.06Rhizopus oryzaeHuang et al (1991)-6.06Rhizopus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzaeHuang et al (1991)31.8-Sludge SolidTien and Huang (19)42.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-15.9Dye-Treated Oil-Palm Fibre1.98-1.98Natural Oil-Palm Fibre28.5G. lucidumNagendra et al. (19)-64.5Treated G. lucidumI.192-10.1Treated A. nigerI.192	
-14.0Amorphous Iron HydroxideMustafa and Haq (1)-9.22Activated CarbonFerro-Gracia et al.0.438-ChitinGonzalez-Davila Millero (1990)-6.89Aspergillus oryzaeHuang et al (1991)-6.06Rhizopus oryzae13.813.8-Aspergillus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzae14.035.7-Sludge SolidTien and Huang (19)42.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19-10.1Treated A. niger10.1	Rahman
-9.22Activated CarbonFerro-Gracia et al.0.438-ChitinGonzalez-Davila Millero (1990)-6.89Aspergillus oryzaeHuang et al (1991)-6.06Rhizopus oryzaeHuang et al (1991)31.8-Aspergillus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzaeHuang et al (1991)35.7-Sludge SolidTien and Huang (1942.9-Chlorella vulgariseAksu et al. (1992)29.0-ZoogloearamigeraI.011.89-Oil-Palm FibresLow et al. (1993)-1.98Natural Oil-Palm Fibre28.5G. lucidumNagendra et al. (19-10.1Treated G. lucidumI.11	
0.438-ChitinGonzalez-Davila Millero (1990)-6.89Aspergillus oryzaeHuang et al (1991)-6.06Rhizopus oryzaeHuang et al (1991)13.8-Aspergillus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzaeHuang et al (1991)35.7-Sludge SolidTien and Huang (1942.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-0il-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19-10.1Treated A. niger10.1	(1988)
-6.89Aspergillus oryzaeHuang et al (1990)-6.06Rhizopus oryzaeHuang et al (1991)-6.06Rhizopus oryzaeHuang et al (1991)31.8-Aspergillus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzaeHuang et al (1991)35.7-Sludge SolidTien and Huang (1942.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-0il-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidum-64.5Treated G. lucidum-10.1Treated A. niger	(1988)
-6.89Aspergillus oryzaeHuang et al (1991)-6.06Rhizopus oryzae13.813.8-Aspergillus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzae16.0635.7-Sludge SolidTien and Huang (19.04)42.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-0il-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidum-64.5Treated G. lucidum-10.1Treated A. niger	and
-6.06Rhizopus oryzae13.8-Aspergillus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzae35.735.7-Sludge SolidTien and Huang (1992)42.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-Oil-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19-64.5Treated G. lucidum-10.1Treated A. niger	
13.8-Aspergillus oryzaeHuang et al (1991)31.8-Treated Aspergillus oryzae35.7-Sludge SolidTien and Huang (1942.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-Oil-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidum-64.5Treated G. lucidum-10.1Treated A. niger	)
31.8-Treated Aspergillus oryzae35.7-Sludge SolidTien and Huang (1942.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-Oil-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19-10.1Treated A. niger	
35.7-Sludge SolidTien and Huang (19)42.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-Oil-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19)-64.5Treated G. lucidum-10.1Treated A. niger	)
42.9-Chlorella vulgariseAksu et al. (1992)29.0-Zoogloearamigera1.89-Oil-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19-64.5Treated G. lucidum-10.1Treated A. niger	
29.0-Zoogloearamigera1.89-Oil-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19-64.5Treated G. lucidum-10.1Treated A. niger	.991)
1.89-Oil-Palm FibresLow et al. (1993)-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19-64.5Treated G. lucidum-10.1Treated A. niger	
-15.9Dye-Treated Oil-Palm Fibre-1.98Natural Oil-Palm Fibre-28.5G. lucidum-64.5Treated G. lucidum-10.1Treated A. niger	
-1.98Natural Oil-Palm Fibre-28.5G. lucidumNagendra et al. (19-64.5Treated G. lucidum-10.1Treated A. niger	
-28.5G. lucidumNagendra et al. (19-64.5Treated G. lucidum-10.1Treated A. niger	
-     64.5     Treated G. lucidum       -     10.1     Treated A. niger	
-     64.5     Treated G. lucidum       -     10.1     Treated A. niger	<del>)</del> 93)
- 10.1 Treated A. niger	
6	
22.2 - Yeast Biomass Brady et al.(1994)	
	Duncan
(1994)	
- 13.5 Banana Pith Low et al (1995)	



-	16.4	Sphagnum Moss Peat	
Table 6 – Credits Adsorpti	•	etals from Waste Streams by	Peat by Yun-Shan Ho, The
	Unive	ersity of Birmingham	

The optimum pH range for copper adsorption was pH 4.0 to 5.0. Since the surface of peat contains acidic functional groups, the optimum pH range is likely to be under weakly acidic conditions. At pH 2.0, there is minimal copper removal, which may be due to the proton exclusion effect, and there is competition of H for surface active sites which lead to minimum or negligible copper ion uptake at low pH, as the pH is increased from 2.0 to 5.0, the removal increases. However, increases above pH 4.0 produce less of an effect than was achieved among 2.0 and 4.0. This effect was most pronounced at low concentrations.

### Adsorption of Nickel

Maximum capacities for adsorption of nickel by different natural adsorbents

Capacity (mg/g)	Xm (mg/g)	Material	Reference
-	7.66	Anaerobically Digested	Gould and Genetelli
		Sludge	(1978)
-	3.40	Delta-Manganese Dioxide	Gray and Malati(1979)
2.40	-	Peat (Rastunsuo)	Tummavuori and Aha
			(1980)
7.45	-	Treated Bagasse	Kumar and Dara (1982)
3.4	-	Treated Acacia Bark	
4.2	-	Treated Laurel Bark	
3.5	-	Treated Techtona Bark	
11.2	-	Eutrophnic Peat	Gosset et al. (1986)
11.7	-	Oligotropic Peat	
5.58	-	Rice Hulls	Suemitsu et al. (1986)
6.16	-	Dyestuff-Treated (Red)	
		Rice Hulls	
6.08	-	Dyestuff-Treated	
		(Yellow) Rice Hulls	
23.0	-	Sphagnum Moss Peat	McLellan and Rock
			(1988)
-	6.75	Amorphous Iron	Mustafa and Haq (1988)
		Hydroxide	
5.22	-	Aspergillus oryzae	Huang et al (1991)
12.4	-	Treated Aspergillus	
		oryzae	
40.8	-	Sludge Solid	Tien and Huang (1991)
-	3.46	China Clay	Sharma et al. (1991)
0.672	-	Sphagnum Peat	Viraraghavan and
			Dronamraju (1993)
0.5	-	Oil-Palm Fibres	Low et al. (1993)
6.46	-	Yeast Biomass	Brady et al. (1994)
-	9.18	Sphagnum Moss Peat	

 Table 7 – Credits Adsorption of heavy metals from waste streams by Peat by Yun-Shan Ho, The University of Birmingham

Comparison of maximum adsorption capacities for lead of various metals with those of peat
---

Capacity (mg/g)	Xm (mg/g)	Material	Reference
-	3.93	Kaolin Clay	Farrah et al. (1980)
-	14.1	Illite Clay	
-	71.8	Montmorillonite Clay	
10.2	-	Treated Bagasse	Kumar and Dara (1982)

Impact Factor value 7.429 | ISO 9001: 2008 Certified Journal Page 630



10.4	-	Treated Acacia Bark	
10.9	-	Treated Laurel Bark	
10.8	-	TeatedTechtona Bark	
0.269	-	Waste Type Rubber	Rowley et al. (1984)
8.90	-	Rice Hulls	Suemitsu et al. (1986)
12.0	-	Dyestuff-Treated (Red) Rice	
		Hulls	
12.0	-	Dyestuff-Treated (Yellow)	
		Rice Hulls	
-	49.9	Moss	Low and Lee (1987)
		(CalymperesdelessertiiBesch)	
-	78.7	Tea Leaves	Tan and Abd. Rahman
			(1988)
-	0.368	Fly-ash	Yadava et al. (1989)
-	1380	Waste Slurry	Srivastava et al. (1989)
19.0	-	Aspergillus oryzae	Huang et al. (1991)
114	-	Treated Aspergillus o	
90.9	-	Sludge Solid	Tien and Huang (1991)
-	0.415	China Clay	Yadava et al. (1991)
-	0.308	Wollastonite	
-	61.8	Sphagnum Peat Moss	Allen et al. (1992)
0.08	-	Oil-Palm Fibres	Low et al. (1993)
-	116	Pencillium Biomass	Niu et al. (1993)
149	-	Titanmium(IV) Oxide	Suzuki et al. (1994)
41.4	-	Yeast Biomass	Brady et al. (1994)
1860	-	Lignin	Srivastava et al. (1994)
-	251	Algae	Ozer et al. (1994)
20.0	-	Peat (Rastunsuo)	Tummavuori and Aho
			(1980)
40.0	-	Sphagnum Moss Peat	McLellan and Rock
			(1988)
-	30.7	Sphagnum Moss Peat	

 Table 8 – Credits adsorption of heavy metals from waste streams by peat by Yun-Shan Ho, The University of Birmingham

### Comparison of adsorption capacities of various adsorbents

Capacity, (mg/g)			Material	Reference		
Cu	Ni	Pb				
5.10	2.40	20.0	Peat (Rastunsuo)	Tummavuori and Aho (1980)		
4.44	7.45	10.2	Treated bagasse	Kumar and Dara (1982)		
3.46	3.4	10.4	Treated Acacia Bark			
3.08	4.2	10.9	Treated Laurel Bark			
3.69	3.5	10.8	Treated Techtona Bark			
3.58	5.58	8.90	Rice Hulls	Suemitsu et al. (1986)		
7.88	6.16	12.0	Dye stuff-Treated (Red) Rice Hulls			
7.0	6.08	12.0	Dye stuff-Treated (Yellow) Rice Hulls			
23.0	-	40.0	Sphagnum Moss Peat	McLellan and Rock (1988)		



14.0	6.75	-	Iron Hydroxide	Mustafa and Haq., (1988)
27.3	-	79.7	Tea Leaves	Tan and Abd. Rahman., (1988)
13.8	5.22	19.0	Aspergillus oryzae	Huang et al., (1991)
31.8	12.4	114	Treated Aspergillus Oryzae	
35.7	40.8	90.9	Sludge Solid	Tien and Huang (1991)
18.5	0.672	-	Sphagnum Peat	Viraraghavan and Dronamraju, (1993)
1.89	0.50	0.08	Oil-Palm Fibres	Low et al. (1993)
22.2	6.46	41.4	Yeast Biomass	Brady et al.(1994)
13.0	9.26	30.2	Sphagnum Moss Peat	

 Table 9 – Credits Adsorption of heavy metals from waste streams by Peat by Yun-Shan Ho, The University of Birmingham

### C. Agricultural wastes

Adsorption capacity of biosorbents obtained from agricultural wastes on the removal of different metal elements

Adsorbents	Metal element	Q (mg/g) or removal	Sources	
		percentage (%)		
Coffee Pulp	Chromium (Cr)	13.48 mg/g	Aguilar et al., 2019	
White yam	Cadmium (Cd)	22.4 mg/g	Asuquo et al., 2018	
Brassica	Nickle (Ni)	1.1 mg/g	Shaikh et al., 2018	
Campestris	Chromium (Cr)	95 mg/g		
waste stem				
Canola Seeds	Lead (Pb)	44.25 mg/g	Affonso et al., 2019	
	Cadmium (Cd)	52.36 mg/g		
Rice Husk	Zinc (Zn)	94.33 %	El Nadi and Abd Alla, 2019	
	Chromium (Cr)	89.20 %	El Nadi and Abd Alla , 2019	
Banana Peel	Copper (Cu)	14.3 mg/g	Thuan et al., 2017	
	Nickle (Ni)	27.4 mg/g		
	Lead (Pb)	34.5 mg/g		
Rice Husk	Chromium (Cr) 97.12 %		Kumar et al, 2017	
Jackfruit Peels	Lead (Pb)	10.1 mg/g	Ibrahim et al.,2020	
	Copper (Cu)	17.5 mg/g		
	Cadmium (Cd)	20.0 mg/g		
	Manganese (Mn)	76.9 mg/g		
	Iron (Fe)	4.40 mg/g		
Pistachio Hull	Nickle (Ni)	14 mg/g	Beidokhti et al., 2019	
Waste				
Ground Nut	Cadmium (Cd)	70.64 %	Vinaykumar et al., 2019	
Shell				
Pongamia		79.9 %		
Pinnata				
Onion Skin		75.45 %		

Table 10 – Credits Journal of Ecological Engineering 2021, 22(3), 249-265



## **D.** A study on heavy metal removal rate: rice husk and fly ash

### Fe removal by different weights of absorbents

The effect of the amount of adsorbent on the removal of Fe ions by rice husk is depicted in Table 3 for varied adsorbent doses of 20, 30, 40, 50 and 60 mg/l. Fe removal using rice husk increased from 68.59% to 99.25% i.e. with the increase of the amount of absorbent concentration , while Fe removal using fly ash varied from 46.18% to 86.757%.

Heavy metal	Adsorbent	In- Fe	Rice husk	Flyash		
	dose	mg/l	Outlet Fe	Removal	Outlet	Removal
			mg/l	ratio %	Fe mg/l	ratio %
Fe	20	11.78	3.7	68.59	6.34	46.18
	30	11.78	2.1	82.17	82.17	58.4
	40	11.78	1.2	89.81	89.81	65.2
	50	11.78	0.09	99.236	99.236	74.788
	60	11.78	0.088	99.253	99.253	86.757

Table 11 - Fe removal efficiency for different absorbent doses

### Pb removal by different weights of absorbents

The effect of the amount of adsorbent on the removal of Pb ions by rice husk is depicted in Table 4 for varied adsorbent doses of 20, 30, 40, 50 and 60 mg/l. Pb removal with rice husk increased from 22.22% to 87.17% i.e. with the increase of the amount of absorbent concentration, while the Pb removal using fly ash varied from 21.79% to 76.06%.

Heavy metal	Adsorbent	In- Pb	Rice husk Flyash			
	dose	mg/l	Outlet Pb mg/l	Removal ratio %	Outlet Pb mg/l	Removal ratio %
Pb	20	1.17	0.91	22.22	0.92	21.79
	30	1.17	0.66	43.59	0.7	40.17
	40	1.17	0.38	67.52	0.46	60.68
	50	1.17	0.28	76.068	0.33	71.795
	60	1.17	0.15	87.179	0.28	76.068

 Table 12 - Pb removal efficiency for different absorbent doses

### Cd removal by different weights of absorbents

The effect of the amount of adsorbent on the removal of Cd ions by rice husk is depicted in Table 5 for varied adsorbent doses of 20, 30, 40, 50 and 60 mg/l. Cd removal using rice husk increased from 26.04% to 67.917% i.e. with the increase of the amount of absorbent concentration, while the Cd removal using fly ash varied from 25.21% to 73.54%.

Heavy metal	Adsorbent	In-	Rice husk	Flyash				
	dose	Cd mg/l	Outlet Cd mg/l	Removal ratio %	Outlet Cd mg/l	Removal ratio %		
Cd	20	0.48	0.36	26.04	0.36	25.21		
	30	0.48	0.31	35.42	0.30	37.50		
	40	0.48	0.24	50.00	0.23	52.08		
	50	0.48	0.19	60.417	0.180	62.500		
	60	0.48	0.154	67.917	0.127	73.542		
Table 13 - Cd removal efficiency for different absorbent doses								

 Table 13 - Cd removal efficiency for different absorbent doses.

### Cu removal by different weights of absorbents

The effect of the amount of adsorbent on the removal of Cu ions by rice husk is depicted in Table 5 for varied adsorbent doses of 20, 30, 40, 50 and 60 mg/l. Cu removal using rice husk increased from 24.49% to 98.177% i.e. with the increase of the amount of absorbent concentration , while Cu removal using fly ash varied from 37.38% to 98.545%.



Heavy metal	Adsorbent	In- Cu	<b>Rice husk</b>		Flyash		
	dose	mg/l	Outlet C	Cu	Removal	Outlet	Removal
			mg/l		ratio %	Cu mg/l	ratio %
Cu	20	5.43	4.10		24.49	3.40	37.38
	30	5.43	2.84		47.70	1.81	66.67
	40	5.43	1.83		66.30	1.01	81.40
	50	5.43	1.210		77.716	0.089	98.361
	60	5.43	0.099		98.177	0.079	98.545

 Table 14 - Cu removal efficiency for different absorbent doses.

### Niremoval efficiency for different absorbent doses

Heavy	Adsorbent	In- Ni	Rice husk	Flyash	Flyash		
metal	dose	mg/l	Outlet Ni	Removal	Outlet	Removal	
			mg/l	ratio %	Ni mg/l	ratio %	
Ni	20	1.74	0.089	94.885	0.095	95.540	
	30	1.74	0.071	95.920	0.085	95.115	
	40	1.74	0.065	96.264	0.076	95.632	
	50	1.74	0.058	96.667	0.070	95.977	
	60	1.74	0.053	96.964	0.069	96.034	

Table 15 - Niremoval efficiency for different absorbent doses

### E. Industrial solid waste

In the case of calcined brick powder

Initial concentration Ci	Equilibrium	qe (mg/g)	Ce/qe() g/l
(mg/l)	concentration Ce (mg/l)		
10	1.863	2.03	0.917
20	4.986	3.75	1.329
30	9.358	5.16	1.810

### Table 16 – Initial pH 2.0

Initial concentration Ci	Equilibrium	qe (mg/g)	Ce/qe() g/l
(mg/l)	concentration Ce (mg/l)		
50	12.23	25.13	0.486
75	28.23	31.18	0.905
100	48.24	34.50	1.398
		34.50	0.2 00

### Table 17 – Ni(II), pH 4.0

Adsorbent used is a solid waste calcined brick powder which is available in large quantities and can be used as an alternative to existing commercial adsorbents for removal of Cr (VI) and Ni (II). The removal of these carcinogenic toxicants was found to depend on dosage, pH, initial concentrations of Cr (VI) and Ni (II) ions and also contact time. The adsorption capacity of CBP adsorbent for Chromium (VI) is more than Nickel (II). Contact time for the maximum adsorption required is 60 min at pH 2.0 for Cr (VI) and 105 min at pH 4.0 for Ni (II). The equilibrium sorption data are satisfactorily fitted with Freundlich and Langmuir equations. The calculated values of the dimensionless separation factor from the Langmuir constant also confirm favourable sorption of Cr (VI) and Ni (II) onto calcined brick powder. Heavy

metal removal with aforesaid CBP adsorbent appears to be technically feasible and eco-friendly too. Also, it helps in reduction of waste generation.

### F. Plastic waste

In the recent times, much research has gone through for identifying the uses of plastic waste as an adsorbent. Additionally, plentiful plastic wastes as adsorbent have been developed and used in numerous environmental amputations

The advantages of the above specified Plastic wastes as adsorbent have been discussed

#### **Polypropylene waste(PP)**



- i. Cost effective for waste water treatment processes
- ii. Can successfully remove copper from waste streams
- iii. Improved oil sorption performance of virgin PP fiber
- iv. Most effective method
- v. Have potential in wastewater treatment

### Low-Density Polyethylene (LDPE) Waste

Reduce production cost of the hydrogel and new method for converting wastes into valuable products

### **Polyethylene Terephthalate (PET) waste**

- i. Effective adsorbent for removing MB and AB25 from aqueous solutions
- ii. Low cost
- iii. Have potentiality of utilized "waste treat waste"
- iv. An effective adsorbent for removal of cadmium ions from aqueous
- v. Low-cost adsorbent
- vi. Good adsorbent to adsorb heavy metal pollutants
- vii. Show better adsorptive properties than activated carbon

### V. FINDINGS AND DISCUSSION

In the comparison of rice husk and flyash

Results showed that low-cost adsorbents can be fruitfully used for the removal of heavy metals with a concentration range of 20–60 mg/l. The results of using real wastewater showed that rice husk was effective in the simultaneous removal of Fe, Pb and Ni, whereas fly ash was effective in the removal of Cd and Cu.It was found that the percentage removal of heavy metals was dependent on the dose of low-cost adsorbent and adsorbent concentration.The contact time necessary for maximum adsorption was found to be two hours.The optimum pH range for heavy metal adsorption was 6–7.0.

### VI. CONCLUSION

As it has been understood that the location of the landfill places a important role in the Leachate ability to contaminate the groundwater. It has to be understood that the landfill with proper implementation of the codalprovisions and suggestions. An appropriate location away from the human living along with proper distance from the waterbodies would be an ideal location.

And while selecting the location other very important parameters such as ground water level, type of soil as well play important role to understand the ability of leachate to percolate though the soil. And in such cases clayey soil would play its role in adsorbing most of the heavy metals compared to other soil types. But however clayey soil could also lead to stagnation in few cases which should be looked into.

Implementing the usage of solid wastes would be a very effective and efficient solution in most of the cases. And some natural adsorbents such as rice husk and milkweed would be effective. And it also be noted that not only a single adsorbent in the site but a combination of two or three adsorbents would be advantageous because a single adsorbent would not be able to adsorb all the different types of heavy metals which would be present in the leachate. In such cases a combination work not only from effectiveness would perspective but also availability point of view, which doesn't require huge quantities of adsorbent at a single point of time in practical scenarios.

And other alternatives are the industrial wastes which could be used as the adsorbent, such as Class IV brick powder, fly ash, blast furnace slag red mud etc which have a very high adsorption rate compared to naturally available adsorbents and could be more viable considering them as industrial wastes.

### REFERENCES

- Acharya, J.Sahu, J. N.Sahoo, B. K.Mohanty, C. R. Meikap B. C. 2009 Removal of chromium (VI) from wastewater by activated carbon developed from Tamarind wood activated with zinc chloride. Chemical Engineering Journal 150 (1), 25–39
- [2]. Adam, F. Kandasamy, K. Balakrishnan, S. 2006 Iron incorporated heterogeneous catalyst from rice husk ash. Journal of Colloid and Interface Science 304 (1), 137 – 143
- [3]. Ajmal, M. Rao, R. A. K. Anwar, S. Ahmad J, Ahmad, R. 2003 Adsorption studies on rice husk: removal and recovery of Cd(II) from wastewater. Bioresource Technology 86 (2), 147–149.
- [4]. Anirudhan, T. S. Sreekumari, S. S. 2011 Adsorptive removal of heavy metal ions from industrial effluents using activated carbon derived from waste coconut buttons. Journal of Environmental Sciences 23(12), 1989–1998.
- [5]. Argun, M. E. Dursun, S. Ozdemir, C. Karatas, M. 2007 Heavy metal adsorption by modified oak sawdust: thermodynamics and kinetics. Journal of Hazardous Materials 141 (1), 77–85.



- [6]. Atieh, M. A. 2011 Removal of chromium (VI) from polluted water using carbon nanotubes supported with activated carbon. Procedia Environmental Sciences4, 281– 293.
- [7]. Bilal, M. Shah, J. A. Ashfaq, T. Gardazi, S. M. H. Tahir, A. A. Pervez, A. Haroon, H. Mahmood 2013 Waste biomass adsorbents for copper removal from industrial wastewater – A review. Journal of Hazardous Materials 263, 322–333.
- [8]. Gupta, V. K. Nayak, A. Agarwal, S.2015 Bioadsorbents for remediation of heavy

metals: current status and their future prospects. Environ. Eng. Res. 20, 1–18.

- [9]. Farajzadeh, M. A. Monji, A. B. 2004 Adsorption characteristics of wheat bran towards heavy metal cations. Sep. Purif. Technol. 38, 197–207.
- [10]. Ibisi, N. E. Asoluka, C. A. 2018 Use of agrowaste (Musa paradisiaca peels) as a sustainable biosorbent for toxic metal ions removal from contaminated water. Chem. Int. 4 52–59.